



## Review Article



# Osteogenesis Imperfecta: A Heterogeneous Heritable Disease

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## Abstract

Bone is a dynamic organ, able to replace old or disrupted tissue through a remodelling process. It contains a relatively small number of cells (osteoblasts, osteocytes, osteoclasts and Mesenchymal Stem Cells (MSCs)) entrenched in a matrix. Perturbation or disruption of the complex molecular pathways controlling MSC proliferation and osteogenic commitment may be determined by mutations affecting key genes in bone development. Osteogenesis Imperfecta (OI) also known as brittle bone disease is a genetic pathology in which bones do not form properly and therefore are fragile and break easily. OI is a heterogeneous congenital heritable disease that mainly affects connective tissues. Nowadays we number 18 types of OI, characterized by various modes of inheritance: autosomal dominant, recessive and X-linked.

## Keywords

Bone; Mesenchymal Stem Cells; Osteogenesis Imperfecta

## Bone Ossification in the Foetus Childhood and Adolescence

Two osteogenic process characterize skeletal development: intramembranous and endochondral ossifications. Ossification is a complex process orchestrated by Mesenchymal Stem Cells (MSCs) able to differentiate either into osteoblasts or chondrocytes [1]. In intramembranous ossification MSCs direct differentiate into osteoblasts and usually occurs in the flat bones as skull, jaw and in the sub-periosteum areas of long bones. Early osteoblasts occurs in cluster called ossification centre, although osteoblasts will be spreaded out by during formation of bone [1]. Osteoblasts secrete osteoid, an uncalcified matrix, which within a few days calcifies trough a mineral salts deposition, thereby entrapping the osteoblasts in the matrix. Entrapped osteoblasts will become osteocytes [2]. As osteoblasts-osteocytes transformation, also osteogenic cells present in the surrounding connective tissue may differentiate into new osteoblasts. Unmineralized bone matrix secreted around the capillaries became a trabecular matrix, while osteoblasts on the surface of the spongy bone form the periosteum. The periosteum represents the protective surface layer above trabecular bone [3,4]. Intramembranous ossification that begins during foetal development prosecutes until adolescence. At birth, the skull and clavicles bones are not fully ossified [4]. Bones plasticity allows skull and shoulders to deform during passage through the birth canal.

In endochondral ossification, process that occurs in long bones formation, mesenchymal stem cells differentiate into chondrocytes and secrete a cartilaginous matrix. Cartilage scaffolds are then replaced by bone, increasing the ability to counteract the compression. For example, six-eight weeks after conception, mesenchymal cells differentiate into chondrocytes which form the cartilaginous skeletal precursor of a long bone [5]. The canonical bone development model involves the differentiation of mesenchymal stem cells into a specific cell lineage fate, depending on the time and signalling [6]. As more matrix is produced, more chondrocytes grow in size. Matrix calcification inhibits nutrients reaching to the chondrocytes. During cartilage grows, capillaries penetrate it, starting the transformation of perichondrium into the bone-producing periosteum. Here, osteoblasts form a periosteal compact bone frame that surround the diaphysis cartilage [7]. From the third month of foetal life, in the periosteal collar bone development creates the primary ossification centre, where ossification begins. Chondrocytes and cartilage continue to grow at the ends of the bone, forming the future epiphyses [4]. While length increases, bone is replacing cartilage in the diaphysis. When the foetal skeleton is fully formed, cartilage remains only at the joint surface and between the diaphysis and epiphysis [4]. After birth, matrix mineralization, chondrocytes' death, invasion of blood vessels from the periosteum, and osteoblasts maturation occurs in the epiphyseal regions, and all these activities centres is referred to as a secondary ossification centre.

Childhood and adolescence are characterized by a progressive growing and by enhanced bone mass. At birth the skeleton weights about 75-90 g; it reaches 2400-3000 g in the young adult [8]. The epiphyseal plate represents the growth area in a long bone. It is a layer of hyaline cartilage where ossification occurs in immature bones. The reserve zone is the region closest to the epiphyseal end of the plate and contains small chondrocytes within the matrix. These cells do not participate in bone growth but secure the epiphyseal plate to the osseous tissue of the epiphysis. The proliferative zone is the next layer towards the diaphysis and contains stacks of slightly larger chondrocytes. It makes new chondrocytes (via mitosis) in order to replace those that die at the diaphyseal end of the plate. Chondrocytes in the next layer, the zone of maturation and hypertrophy, are older and larger than those in the proliferative zone; the more mature cells are located closer to the diaphyseal end of the plate [8]. Bone longitudinal growth is a result of cellular division in the proliferative zone and cellular maturation within the zone of maturation and hypertrophy, respectively. In calcified matrix, the zone closest to the diaphysis, most chondrocytes are dead. Capillaries and osteoblasts from the diaphysis penetrate this zone; osteoblasts secrete bone tissue on the remaining calcified cartilage. Thus, the zone of calcified matrix connects

the epiphyseal plate to the diaphysis. A bone grows in length when osseous tissue is added to the diaphysis. The growth rate is controlled by hormones; bones continue to grow in length until early adulthood. Bone growth includes also increasing in diameter, this can continue even after longitudinal growth ceases and it is called appositional growth. Osteoclasts resorb old bone that lines the medullary cavity, while osteoblasts, via intramembranous ossification, produce new bone tissue beneath the periosteum. The canonical endochondral ossification leads to the apoptosis of hypertrophic chondrocytes and the vascular invasion that induce the osteoclast precursors to remove cartilage, while the osteoblast begin to form new bone [6]. Recently have been reported the ability of chondrocyte to transdifferentiate into osteoblast during endochondral ossification process. Three majors mechanistically different transdifferentiation process has been described. The first two process involve an intermediate transdifferentiation, while the third model direct transdifferentiation occur [6]. The chondrocyte switch into Osteogenic Precursor (OP) suggests that in the growth plate immature chondrocytes may differentiate into a transient osteogenic precursor in the metaphysis [9]. In the second intermediate transdifferentiation, chondrocytes dedifferentiate into immature chondrocytes and to redifferentiate into osteogenic lineage. It has been described during embryonic and post-natal development [9]. The third model, described as direct transdifferentiation, is the method by which post-natal secondary ossification occurs. Mature, hypertrophic and not apoptotic chondrocytes differentiate directly into osteoblasts and then into osteocytes [6].

## Osteogenesis Imperfecta: History

Perturbation or disruption of the molecular pathways controlling MSC proliferation and osteogenic commitment may be due to mutations in key genes in bone development. Osteogenesis Imperfecta (OI) also known as brittle bone disease is a genetic pathology in which bones do not form properly and therefore are fragile and break easily [10]. It is a heterogeneous heritable disease that mainly affects connective tissues. The estimated incidence is approximately 1 per 20,000 live births [9]. The various genetic mutations that cause OI in 85% of cases affect type I collagen- one of the critical components of bone matrix, either quantitatively or qualitatively [11]. The first studies on OI were done in 1788 by Olof Jakob Ekman; it can actually be considered a very ancient pathology. During archaeological studies some Egyptian mummies were found; the description of their skull, teeth and flat bones abnormalities suggests that they were affected by OI [12]. In 1979 Sillence et al., described OI as a heterogeneous disease and, for the first time, they proposed a classification of at least four distinct types [13].

## Phenotypes of Osteogenesis Imperfecta

The hallmarks of the brittle bone dysplasia Osteogenesis Imperfecta are skeletal deformities and bone fragility that causes low bone mass and bone fractures [13]. The severity of the disease can range from mild to lethally severe. Frequent and multiple fractures typically lead to bone deformities and short stature. In the mildest forms, fractures tend to decrease in the adults, but postpartum or in menopause may re-occurs [14]. Since type I collagen is such an important structural protein in many connective tissues, people with OI may also experience fragile skin, weak muscles, loose joints, easy bruising, frequent nosebleeds, brittle teeth, blue sclerae, and hearing loss. The clinical features can be divided into skeletal and extra-skeletal manifestations. Skeletal features involve excess or atypical fractures, short stature, scoliosis, and basilar skull deformities, while extra-skeletal symptoms include hearing loss, it is found in 50% of adults by age 50 and in 5% of children with OI [15]. Dentinogenesis Imperfecta (DI), consist in small deformed teeth, which present opalescent and opaque dentin. Malocclusion and DI, are the main dental abnormalities which may occur. The phenotype is variable also within the same patient, with some teeth appearing normal and others

being affected [16]. The sclerae may be blue or grey, colour can be stable or became less dark. Blue sclerae it is characteristic of OI type I and of mildest forms [17]. Connective tissue abnormalities may result in dislocation of head of the radius and the joint. 36% of patients that manifest hypercalciuria, may result in renal calculi [18,19]. Cardiovascular complications leads to aortic root dilatation and mitral valve prolapse. Also, neurological manifestations have been reported and include macrocephaly, hydrocephalus, basilar invagination and cervical spine kyphosis. Hearing impairment is a common symptom in OI, from 39% to 57.9% [12]. Its prevalence increases over the time, it usually manifests between the second and fourth decade, and it is progressive [20]. Patients with OI are characterized by low areal Bone Mineral Density (aBMD), associated to lower bone size and lower volumetric BMD [21]. Histomorphometric evaluation shows increased cortical porosity, low cortical width and trabecular bone volume reduction [22,23]. However, bone may appear hypermineralized with smaller and abundant mineral crystals, process associated with lower mechanical strength [24]. In Figure 1 are reported some examples of AD OI types (A, B, C), AR that affects bone mineralization (D, E) and also few cases of abnormal collagen post-translational modification (E, F).



**Figure 1:** Clinical features associated with Osteogenesis Imperfecta. In a blue box some examples of dominant forms of OI, part A type I OI, part B a radiographic images showing severe curvature of the legs in type II OI, and part c a picture of type III OI patients. In a red box, part D illustration the radiograph of limbs in type VI OI, part E the radiographs of the patient's limbs showing fractures and interosseous membrane calcifications development as well as the initial evidence of periosteal calcification (arrows) Radial head dislocation is visible on the forearm anteroposterior view patients with Osteogenesis Imperfecta. In a green box part F a patient at 5 years affected by type VII OI; the patient has a normal head, white sclerae, small thorax and shortening of the proximal segment of the upper and lower extremities. In the left, detail of arm and legs showing osteopenia, undertubulation and severe deformities, consistent with a severe deforming form of Osteogenesis Imperfecta. Part F patient affected by type VIII OI, radiograph show bowed limbs and severe osteopenia and undertubulated long bones [25-28].

## Genetic Classification and Pathophysiology of OI

Mutations in genes coding for the  $\alpha 1$  and  $\alpha 2$  chains of the heterotrimeric type I collagen [(1)2 2(1)] were associated to Osteogenesis imperfecta in 1980 [15]. Type I collagen is the most abundant protein of bone, skin, and tendon; in teeth and bone it plays a fundamental role in the mineralization process. COL1A1 and COL1A2 mutations are the most common defects (85% of OI cases) transmitted as autosomal inheritance. COL1A1 loss-of-function mutations cause a quantitative loss of in  $\alpha 1(I)$  chains of type I collagen trimers; they typically generate a mild form of OI (type I) [15]. Quantitative defects are associated with the milder osteogenesis imperfecta type I. Other mutations in COL1A1 or COL1A2, such as glycine substitutions in the Gly-X-Y repeat, lead to structural defects of the collagen triple helix. They exert a dominant negative effect on the normal collagen chains upon trimers formation, and result in either moderate, severe or lethal OI. Structural collagen defects can cause moderate and progressive deforming type IV and type III, respectively, as well as lethal type II [29].

In the past decade, a wide variety of genes encoding proteins involved in type I collagen synthesis, processing, secretion and post-translational modification, respectively, as well as in genes coding for proteins that regulate the differentiation and activity of bone-forming cells have been shown to cause Osteogenesis Imperfecta [30]. In 2006 CTRAP, was identified as the first gene causing recessively inherited OI, opening the way to an exciting new information about the genetics and mechanism

of this bone dysplasia.

OI classification initially included four phenotypes (Sillence classification [31]): type I, or the non-deforming type, autosomal dominant mild form, characterized by blue sclerae, type II, with autosomal recessive inheritance the most severe form (lethal perinatally) an type III, autosomal recessive inheritance severe form, that manifests progressive deformity and type IV autosomal dominant inheritance form with moderate severity [14]. Clinical manifestation of COL1A1 and COL1A2 mutations are reported in box1 [32] (Table 1).

OI type	Phenotype
I/I	Mild, nondeforming and lethal forms
2/II, VII, VIII, IX, XII	Severe, seen as perinatal
3/III, VII, VIII, IX, XI, X, VI, XV	Moderate to severe, progressively deforming
4/IV, VII, XI, XIII, VI, XV, XIV	Moderate
5/V	Moderate with calcification of the interosseous membrane and/or hypertrophic callus

**Table 1:** The International Nomenclature Group for Constitutional Disorders ICHG of the Skeleton 2009.

The discovery of new OI associated genes has led to two new approaches to classification: 1) a more clinically based approach in which the new recessive types are included under the Sillence-types, 2) a genetic–functional approach in which the Sillence types I–IV are reserved for mutations in COL1A1 or COL1A2 and new genes are given additional type numbers based on the mutation without clinical correlation (Table 2)

Mutated gene	Encoded protein	Inheritance	Localization	Severity	OI type
Impairment of collagen synthesis and structure					
COL1A1	Collagen $\alpha 1$	AD	matrix structural component	Mild to lethal	I,II,III or IV
COL1A2	Collagen $\alpha 2$	AD	matrix structural component	Moderate to lethal	I,II,III or IV
Compromised bone mineralization					
IFITM5	BRIL	AD	bone-restricted interferon-induced transmembrane like protein	variable severity	V
SERPINF1	PEDF	AR	collagen-binding protein/pigment epithelium derived factor	Moderate to severe	VI
Abnormal collagen post-translational modification					
CTRP	CTRAP	AR	endoplasmic reticulum	Severe to lethal	VII
P3H1 (LEPRE1)	P3H1	AR	endoplasmic reticulum	Severe to lethal	VIII
PPIB	PPIase B	AR	endoplasmic reticulum	Moderate to severe	IX
Compromised collagen processing and crosslinking					
SERPINH1	HSP47	AR	endoplasmic reticulum-golgi	Severe to lethal	X
FKBP10	FKBP65	AR	endoplasmic reticulum	Moderate to severe	XI
PLOD2	LH2	AR	endoplasmic reticulum	Moderate to severe	no type
BMP1	BMP1	AR	endoplasmic reticulum	Moderate to severe	XII
Alternate osteoblast differentiation and function					
SP7	Transcription factor SP7/ osterix	AR	nucleus	Mild to moderate	XIII
TMEM38B	TRIC-B	AR	cation channel	Moderate to severe	XIV
WNT1	WNT1	AR/AD	secreted signal molecule	Moderate to severe	XV
CREB3L1	OASIS	AR	endoplasmic reticulum-golgi	Severe	XVI
SPARC	SPARC/osteonectin	AR	matrix	Moderate to severe	XVII
MBTPS2	S2P	XLR	endoplasmic reticulum-golgi	Moderate to severe	XVIII

**Table 2:** Genetic classification of OI.

[31]. In 2009, the International Nomenclature Group for Constitutional Disorders ICHG of the Skeleton (INCDS) proposed a new classification, divided in five different groups based on phenotypical traits. The individual OI disorders still retain their Roman identification number and in addition they are classified with an Arabic numeral that indicates the phenotypic description, see table 1. The large number of causative genes discovered since 2006 has complicated the classic classification of the disease, and, although a new genetic classification system is widely used, it is still debated. In OI patients have been described a huge defects in proteins with very different functions, ranging from structural to enzymatic and from intracellular transport to chaperones [10]. Nowadays we number 18 different types of OI, distinguished by autosomal dominant, recessive and X-linked inheritance.

### Type I

Caused to COL1A1 or COL1A2 mutation. The clinical manifestations include osteopenia, vertebral fractures, usually not deforming but can lead to scoliosis. Fractures are mostly during childhood and it is associated with extraskeletal defects as blue sclera, presenile deafness, and aortic regurgitation. At X-ray skull and spinal cord shows thin cortices [11].

### Type II

Fractures manifests even before birth and affects rib, long bone, and skeletal fractures. Due to respiratory insufficiency, central nervous system malformations, and hemorrhages death can occur. Other phenotypical traits manifested are blue sclera and dentinogenesis imperfecta. X-ray showed undermineralization, severely deformed extremities [11].

### Type III

Fractures can start in utero or at birth, bringing to a progressive deformity and scoliosis. At birth they present triangular facies, blue sclera and manifests dentinogenesis imperfecta. Are characterized by short stature, severe long bone deformities, fractures, and possibility to develop respiratory hypoplasia. Frequently manifests hearing loss [11].

### Type IV

At birth they may present progressive deformity. Typical traits of type IV concerns greyish or white sclera, dentinogenesis imperfecta, short stature, may have long bone bowing, scoliosis, and joint laxity, although phenotypes are significantly variable [11].

### Type V

They present fractures and hypertrophic calluses that is associated with progressive deformity. Calcification of the interosseous membranes of the forearm can lead to decreased hand mobility and radial head dislocation, moreover

present irregular mesh-like bone. X-ray investigation shows macrocephaly and Wormian bones, scoliosis [11].

### Type VI

Phenotype appear from moderate-to-severe deformities. They may have blue sclera. At birth are healthy with subsequent progressively severe deformities. It is characterized by unmineralized bone and “fishscale” pattern on iliac crest biopsies. Back reveals scoliosis and compression fractures [11].

### Type VII

Phenotype of type VII OI (CRTAP deficiency) overlaps Sillence types II and III but has distinctive features. Causes severe to lethal osteochondrodysplasia with rhizomelia, neonatal fractures, broad undertubulated long bones, frail ribs. Sclerae are white or light grey. Severe growth deficiency and “popcorn” calcifications of epiphyses are seen in individuals who survive into childhood, which are also observed in about half of type III OI patients [11].

### Type VIII

They manifest progressive deformities and short stature. Back reveals severe scoliosis and could be similar to OI Type II/III. Individuals with types VII and VIII OI exhibit a similar phenotype due to the mutual stabilization between CRTAP and P3H1 [11].

### Type IX

Blue sclera and severe deformities are the common traits. They may have short stature. Back reveals kyphoscoliosis and skeletal features similar to OI Type II/III/IV [11].

### Type X

It is characterized by severe bone deformities and multiple fractures, generalized osteopenia, dentinogenesis imperfecta, and blue sclera. They may have renal stones [11].

### Type XI

Severe deformities are seen. Patients with type OI type XI have severe progressive deformation and may have joint contractures, dentinogenesis is normal [11].

### Type XII

They have recurrent fractures and mild bone deformities, delayed tooth eruption, generalized osteoporosis, delayed eruption of teeth, absence of dentinogenesis imperfecta, normal hearing, and white sclera [11].

### Type XIII

Fractures manifests after birth recurrently, hearing is normal and sclerae are white. They present a mild bone deformity, and

delayed tooth eruption, but they do not have dentinogenesis imperfecta. Radiographic examination of the skeletal system revealed bowing of the upper and lower limbs [11].

#### Type XIV

Characterized by huge degrees of severity with multiple fractures, that occurs prenatally or at approximately 6 years of age. Osteopenia, absence of dentinogenesis imperfecta, normal sclera, and hearing have been reported [33].

#### Type XV

Phenotype variate from severity, ranging from mild to progressively deforming, which can occasionally lead to early infant death. They present short stature, no joint or skin hyperlaxity, sclera are white, and teeth appeared normal. Lumbar spine areal bone mineral density was very low [34].

#### Type XVI

Fractures can occur in uterus, at birth manifest short stature, multiple fractures, and soft calvarial bones and widely open fontanelles. During adolescence and also adults present blue sclerae, teeth are normal and progressive hearing loss [35].

#### Type XVII

No skeletal abnormalities were reported at birth. After birth manifest first fractures and may present multiple vertebral compression fractures of the thoracic spine and kyphoscoliosis. Sclerae are white and dentinogenesis are normal but they may have mild joint hyperlaxity [36].

#### Type XVIII

Prenatal fractures of ribs and long bones can occur. The present moderate short stature, blue sclerae, pectus carinatum, bowing of lower extremity long bones, variable scoliosis, chest deformity, striking tibial anterior angulation and generalized osteopenia [37].

## Multomics Analysis

Osteogenesis Imperfecta had been known since the early 1980s as a disease caused by mutations in either of the genes encoding type I collagen (COL1A1 and COL1A2). Since 2006, new mutations in collagen-related genes with different inheritance patterns have been found to cause OI [38]. During these 25 years' time lapse, diagnostic and research techniques have been improved. In 2001, the Human Genome Project (HGP) revealed for the first time the entire sequence of the human genome. This achievement has allowed a big work progress in biomedical research. Moreover, since 2007 the technical advancement of Next Generation Sequencing (NGS) has completely revolutionized the approach to DNA sequencing [39]. Through NGS, scientists can compare a large number

of different genomes: this paved the way to many population studies. Furthermore, NGS makes it possible to compare the genome of a healthy individual with the genome of an affected individual and to find out gene(s) associated with a specific disease [40].

The NGS revolution applied to OI, combined with the traditional approaches, has unravelled since 2006 it's astonishing genetic heterogeneity: fifteen novel disease loci have been discovered in ten years' time [41]. Since 2006, seven of the disease genes discovered, cause AR forms of OI, encoding for proteins which are involved in collagen I modifications, processing, folding, cross-linking. Eight additional genes, whose defects cause either AR and AD forms, code for proteins involved in several osteoblast functions and survival. Moreover, epigenetic modifications of DNA (i.e., Cytosine methylation) it has been described as associated to *de novo* OI causing mutations [26], and contribute to explain recurrent *de novo* OI mutations. Scientific progress has prompted a few neologisms ending with -OMICS that refer to various study fields in biology, such as genomics, transcriptomics, proteomics, metabolomics. The related suffix -ome is used to address the study objects of such fields, such as the genome, transcriptome, proteome or metabolome, respectively. Genomics is an interdisciplinary research field focusing on the structure, function, evolution, mapping, and editing of genomes. A genome is a complete set of DNA, including all the genes of an individual. In contrast to genetics, which refers to the study of single genes and their roles in inheritance, genomics aims at the collective characterization and quantification of genes. Likewise, transcriptome refers to all RNA molecules in one cell or a cell population. Unlike DNA, which is the same in every somatic cell of an organism, RNA molecules reflect the expression profile of a specific cell type. Accordingly, proteomics and metabolomics represent the set of proteins and metabolites, respectively. Proteomics is used to quantify peptides abundance, modification, and interaction. Metabolomics simultaneously quantifies multiple small molecules, such as amino acids, fatty acids, carbohydrates, or other products of cellular metabolic functions. Metabolite levels and relative ratios reflect metabolic function, and out-of-normal range perturbations are often indicative of disease. Quantitative metabolite levels allowed the discovery of previously unknown genetic loci regulating small molecules, or their relative ratios, in plasma and other tissues [42,43]. Associated technologies include Mass Spectrometry (MS)-based approaches to quantify both relative and targeted small molecule abundances [44,45].

The omics field has been driven largely by technological advances that have made cost-efficient, high-throughput analysis of biologic molecules possible. Each type of omics data, on its own, typically provides a list of differences associated with the disease or with the population studied [46]. Novel and unexpected OI candidate genes have been identified since 2010 by means of -omics approaches, in table 3 an overview

about of OI-related genes identification from 1983 to 2016 [41]. Nevertheless, new OI disease genes are to be discovered, particularly in rare recessive forms which occur in inbred pedigrees not yet characterized at the molecular level. The candidate genes list is expected to grow in the near future, thanks to the combined -omics approaches.

Patients with OI type V are characterized by Calcification in Interosseous Membranes [49].

Unfortunately, there is no known therapy for the cure of osteogenesis imperfecta and the goal of treatment is addressed to prevent the skeletal deformities. As bone fragility is the most important feature of OI, the treatment aims to reduce

Defective gene	Year	Methodological approach	OI type	Inheritance
COL1A1	1983	candidate gene	I,II,III,IV	AD
COL1A2	1984	candidate gene	I,II,III,IV	AD
IFITM5	2012	Wes, homozygosity mapping+targed NGS	V	AR
SERPINF1	2011	Wes, gwla+targed NGS	VI	AR
CTRP	2006	gwla in inbred families+candidate gene	VII	AR
P3H1 (LEPRE1)	2007	candidate gene	VIII	AR
PPIB	2009	candidate gene	IX	AR
SERPINH1	2010	candidate gene	X	AR
FKBP10	2010	homozygosity mapping+targed NGS	XI	AR
PLOD2	2012	homozygosity mapping+candidate gene	no type	AR
BMP1	2012	homozygosity mapping+candidate gene	XII	AR
SP7	2010	homozygosity mapping+candidate gene	XIII	AR
TMEM38B	2012	autozygosity mapping+wes	XIV	AR
WNT1	2013	wes+gwla+target ngs	XV	AR
CREB3L1	2013	candidate gene	XVI	AR
SPARC	2015	wes	XVII	AR
MBTPS2	2016	gwla+exome sequencing	XVIII	AR

**Table 3:** Years and method to identify the OI-related genes. Wes (Whole Exome Sequencing); gwla (Whole Genome Linkage Analysis); nsg (Next Generation Sequencing).

## Diagnosis and Treatment

The fractures are the most important clinical features occurring in OI, even if 10% of OI patients have not reported fractures in long bone during childhood. Many patients develop the osteoporosis and levels of serum and urine markers of bone turnover (such as osteocalcin, alkaline phosphatase and amino-terminal telopeptide of type 1 collagen) associated to histomorphometry analyses help to perform OI diagnosis.

Often OI patients show blueness of the sclera, hearing loss, dentinogenesis imperfect, short stature, joint hypermobility [47]. In adult patients, cardiovascular complications may occur [48]. In particular, patients with OI type 1 is characterized by bone loss, blue-gray sclera and susceptibility to hearing loss in adolescence and young adult life while bone deformities is less frequent [47]. OI type 2 is extremely severe. Most fetus with OI type 2 are diagnosed prenatally and termination of pregnancy often occurs [47]. Patients with OI type 3 are diagnosed during the childhood and they show bone fragility and multiple fractures causing skeletal deformity. At the birth the sclerae may be blue and, generally, the sclerae become less blue with age [47]. Patients with OI type IV have osteoporosis and recurrent fractures with deformity of long bones [47].

osteoclastic activity. Therefore, bisphosphonates are considered the gold standard treatment for children with moderate to severe OI [50]. Immunotherapy, such as with Antisclerostin antibody, Cathepsin K antibody or anti-RANK-ligand antibody, has been proposed for improving bone mineral density in OI patients [51].

New therapeutic approaches are then required against OI. A challenge to counteract this genetic disease could be represented by the cell-based therapy based on the possibility of mesenchymal stem cells transplantation from health donors to OI patients. Preclinical studies have been conducted in OI murine models [52,53] and clinical attempts have been performed using whole bone marrow or fractionated MSCs in OI children [54,55] and a reduced number of fractures in these children have been reported by the authors [55]. However, further investigations need to be addressed to consider efficacy the cell-based therapy and new technologies such as CRISPR/Cas system, TALEN, and zinc-finger nucleases for genetic correction may be considered an important challenge to correct the DNA in adult cells.

## Natural Evolution of OI

Musculoskeletal disorders and hearing issue are the most important organ compromised in adult OI. In a study

conducted in 37 patients, Sillence reported that foot pain, hearing deficit, and back pain are the most diffuse physical problems in OI patients [56]. The physical status of PI patients is usually evaluated on the basis of the ability to ambulate, the presence of fractures as well as the presence of deformities such as scoliosis or bowing [57,58].

The lungs are often compromised as the chest walls are greatly affected causing airway obstruction, hypertension in the pulmonary district and sleep apnea [59]. Scoliosis and pulmonary affections in OI patients are strongly correlated with reduced physical activity [60]. This finding is important as OI patients are sedentary and have an elevated BMI [61]. In severe OI respiratory diseases induce the death of the patients [61,62]. Dentinogenesis imperfect affects up to 50% of the OI patients [59]. Therefore, OI patients need frequent dental attentions.

Importantly, physical limitations influence the life quality of OI patients with consequences in the social field, self-image and independence. Interestingly, in a study conducted by administrating the questionnaires, the authors found that OI patients, although significant physical limitations, have high social achievement and employment. In addition, the authors reported that the inability to ambulate correlated with an elevate rate of unemployment [60].

## Conclusion

Osteogenesis Imperfecta is a disorder arising from a large spectrum of genetic mutations. In the last years new genetic causes have been identified in OI disease even if most cases of OI are due to COL1A1/ A2 mutations. Therefore, further clinical investigations finalized to improve therapeutic approaches against the new genetic discoveries are needed.

Bone deformities and collagen defects induce important pathological conditions affecting various internal organs. There is not a known definitive cure for OI, and therapeutic treatment aim to prevent or reducing the symptoms. Cell transplantation as well as gene therapy could represent important challenges to counteract severe OI. However, these therapeutic approaches need further investigations.

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## Conflicts of Interest

The authors declare no conflict of interest.

## References

- Dalle Carbonare L, Innamorati G, Valenti MT (2012) Transcription Factor Runx2 and its Application to Bone Tissue Engineering. *Stem Cell Rev* 8: 891-897.
- Valenti MT, Dalle Carbonare L, Mottes M (2017) Osteogenic Differentiation in Healthy and Pathological Conditions. *Int J Mol Sci* 18: 41.
- Atkins GJ, Findlay DM (2012) Osteocyte regulation of bone mineral: a little give and take. *Osteoporos Int* 23: 2067-2079.
- Berendsen AD, Olsen BR (2015) Bone development. *Bone* 80: 14-18.
- Blair HC, Larrouette QC, Li Y, Lin H, Beer-Stoltz D, et al. (2017) Osteoblast Differentiation and Bone Matrix Formation *In Vivo* and *In Vitro*. *Tissue Eng Part B Rev* 23: 268-280.
- Aghajanian P, Mohan S (2018) The art of building bone: emerging role of chondrocyte-to-osteoblast transdifferentiation in endochondral ossification. *Bone Res* 6: 19.
- Dalle Carbonare L, Valenti MT, Bertoldo F, Zanatta M, Zenari S, et al. (2005) Bone microarchitecture evaluated by histomorphometry. *Micron* 36: 609-616.
- Bailey DA, McKay HA, Mirwald RL, Crocker PR, Faulkner RA (1999) A six-year longitudinal study of the relationship of physical activity to bone mineral accrual in growing children: the university of Saskatchewan bone mineral accrual study. *J Bone Miner Res* 14: 1672-1679.
- Ono N, Ono W, Nagasawa T, Kronenberg HM (2014) A subset of chondrogenic cells provides early mesenchymal progenitors in growing bones. *Nat Cell Biol* 16: 1157-1167.
- Ben Amor M, Rauch F, Monti E, Antoniazzi F (2013) Osteogenesis imperfecta. *Pediatr Endocrinol Rev* 10: 397-405.
- Sam JE, Dharmalingam M (2017) Osteogenesis Imperfecta. *Indian J Endocrinol Metab* 21: 903-908.
- Kuurila K, Kaitila I, Johansson R, Grénman R (2002) Hearing loss in Finnish adults with osteogenesis imperfecta: a nationwide survey. *Ann Otol Rhinol Laryngol* 111: 939-946.
- Sillence DO, Senn A, Danks DM (1979) Genetic heterogeneity in osteogenesis imperfecta. *J Med Genet* 16: 101-116.
- Tournis S, Dede AD (2018) Osteogenesis imperfecta - A clinical update. *Metabolism* 80: 27-37.
- Forlino A, Marini JC (2016) Osteogenesis imperfecta. *Lancet* 387: 1657-1671.
- Sillence D, Butler B, Latham M, Barlow K (1993) Natural history of blue sclerae in osteogenesis imperfecta. *Am J Med Genet* 45: 183-186.
- Barron MJ, Sinead T, MacKie I, Michael JD (2008) Hereditary dentine disorders: dentinogenesis imperfecta and dentine dysplasia. *Orphanet J Rare Dis* 3: 31.
- Glorieux FH, Ward LM, Rauch F, Lalic L, Roughley PJ, et al. (2002) Osteogenesis imperfecta type VI: a form of brittle bone disease with a mineralization defect. *J Bone Miner Res* 17: 30-38.
- Horwitz EM, Darwin JP, Lorraine AF, Winston K, Jeffrey CM, et al. (1999) Osteogenesis imperfecta calls for caution - Reply. *Nat Med* 5: 466-467.

20. Swinnen FK, Paul JC, Anne MP, Sofie S, Fransiska M, et al. (2011) Osteogenesis Imperfecta: the audiological phenotype lacks correlation with the genotype. *Orphanet J Rare Dis* 6: 88.
21. Folkestad L, Hald JD, Hansen S, Gram J, Langdahl B, et al. (2012) Bone geometry, density, and microarchitecture in the distal radius and tibia in adults with osteogenesis imperfecta type I assessed by high-resolution pQCT. *J Bone Miner Res* 27: 1405-1412.
22. Rauch F, Travers R, Parfitt AM, Francis HG (2000) Static and dynamic bone histomorphometry in children with osteogenesis imperfecta. *Bone* 26: 581-589.
23. Albert C, Jameson J, Smith P, Harris G (2014) Reduced diaphyseal strength associated with high intracortical vascular porosity within long bones of children with osteogenesis imperfecta. *Bone* 66: 121-130.
24. Imbert L, Aurégan JC, Pernelle K, Hoc T (2014) Mechanical and mineral properties of osteogenesis imperfecta human bones at the tissue level. *Bone* 65: 18-24.
25. Mottes M, Gomez Lira MM, Valli M, Scarano G, Lonardo F, et al. (1993) Paternal mosaicism for a COL1A1 dominant mutation (alpha 1 Ser-415) causes recurrent osteogenesis imperfecta. *Hum Mutat* 2: 196-204.
26. Corradi M, Monti E, Venturi G, Gandini A, Mottes M, et al. (2014) The recurrent causal mutation for osteogenesis imperfecta type V occurs at a highly methylated CpG dinucleotide within the IFITM5 gene. *J Pediatr Genet* 3: 35-39.
27. Venturi G, Gandini A, Monti E, Dalle Carbonare L, Corradi M, et al. (2012) Lack of expression of SERPINF1, the gene coding for pigment epithelium-derived factor, causes progressively deforming osteogenesis imperfecta with normal type I collagen. *Journal of Bone and Mineral Research* 27: 723-728.
28. Valli M, Barnes AM, Gallanti A, Cabral WA, Viglio S, et al. (2012) Deficiency of CRTAP in non-lethal recessive osteogenesis imperfecta reduces collagen deposition into matrix. *Clin Genet* 82: 453-459.
29. Saga S, Nagata K, Chen WT, Yamada KM (1987) pH-dependent function, purification, and intracellular location of a major collagen-binding glycoprotein. *J Cell Biol* 105: 517-527.
30. Tedeschi E, Antoniazzi F, Venturi G, Zamboni G, Tatò L (2006) Osteogenesis imperfecta and its molecular diagnosis by determination of mutations of type I collagen genes. *Pediatr Endocrinol Rev* 4: 40-46.
31. Kliegman R, Nelson WE (2011) *Nelson textbook of pediatrics*. Elsevier/Saunders, Philadelphia PA, USA.
32. Marini JC, Forlino A, Bächinger HP, Bishop NJ, Byers PH, et al. (2017) Osteogenesis imperfecta. *Nat Rev Dis Primers* 3: 17052.
33. Shaheen R, Alazami AM, Alshammari MJ, Faqeih E, Alhashmi N, et al. (2012) Study of autosomal recessive osteogenesis imperfecta in Arabia reveals a novel locus defined by TMEM38B mutation. *J Med Genet* 49: 630-635.
34. Fahiminiya S, Majewski J, Mort J, Moffatt P, Glorieux FH, et al. (2013) Mutations in WNT1 are a cause of osteogenesis imperfecta. *J Med Genet* 50: 345-348.
35. Symoens S, Fransiska M, Sanne D, Callewaert B, Dheedene A, et al. (2013) Deficiency for the ER-stress transducer OASIS causes severe recessive osteogenesis imperfecta in humans. *Orphanet J Rare Dis* 8: 154.
36. Rosset EM, Bradshaw AD (2016) SPARC/osteonectin in mineralized tissue. *Matrix Biol* 52-54: 78-87.
37. Lindert U, Cabral WA, Ausavarat S, Tongkobpetch S, Ludin K, et al. (2016) MBTPS2 mutations cause defective regulated intramembrane proteolysis in X-linked osteogenesis imperfecta. *Nat Commun* 7: 11920.
38. Marini JC, Blissett AR (2013) New genes in bone development: what's new in osteogenesis imperfecta. *J Clin Endocrinol Metab* 98: 3095-103.
39. Metzker ML (2010) Sequencing technologies - the next generation. *Nat Rev Genet* 11: 31-46.
40. Rizzo JM, Buck MJ (2012) Key principles and clinical applications of "next-generation" DNA sequencing. *Cancer Prev Res* 5: 887-900.
41. Mottes M, Giovanni M (2017) Hunting Novel Disease Genes in the Next Generation Sequencing Era: Lessons from Osteogenesis Imperfecta. *J of Genetics and genomes*.
42. Ghazalpour A, Brian JB, Diana S, Che N, Orozco L, et al. (2014) Genetic regulation of mouse liver metabolite levels. *Mol Syst Biol* 10: 730.
43. Gieger C, Geistlinger L, Altmaier E, de Angelis MH, Kronenberg F, et al. (2008) Genetics meets metabolomics: a genome-wide association study of metabolite profiles in human serum. *PLoS Genet* 4: e1000282.
44. Dettmer K, Aronov PA, Hammock BD (2007) Mass spectrometry-based metabolomics. *Mass Spectrom Rev* 26: 51-78.
45. Dunn WB, Broadhurst DI, Atherton HJ, Goodacre R, Griffin JL (2011) Systems level studies of mammalian metabolomes: the roles of mass spectrometry and nuclear magnetic resonance spectroscopy. *Chem Soc Rev* 40: 387-426.
46. Hasin Y, Seldin M, Lusic A (2017) Multi-omics approaches to disease. *Genome Biol* 18: 83.
47. Van Dijk FS, Sillence DO (2014) Osteogenesis imperfecta: clinical diagnosis, nomenclature and severity assessment. *Am J Med Genet A* 164: 1470-1481.
48. Radunovic Z, Wekre LL, Diep LM, Steine K (2011) Cardiovascular abnormalities in adults with osteogenesis imperfecta. *Am Heart* 161: 523-529.
49. Glorieux FH, Rauch F, Plotkin H, Ward L, Travers R, et al. (2000) Type V osteogenesis imperfecta: a new form of brittle bone disease. *J Bone Miner Res* 15: 1650-1658.
50. Bishop N, Adami S, Ahmed SF, Antón J, Arundel P, et al. (2013) Risedronate in children with osteogenesis imperfecta: a randomised, double-blind, placebo-controlled trial. *Lancet* 382: 1424-1432.
51. Glorieux FH, Devogelaer JP, Durigova M, Goemaere S, Hemsley S, et al. (2017) BPS804 Anti-Sclerostin Antibody in

- Adults With Moderate Osteogenesis Imperfecta: Results of a Randomized Phase 2a Trial. *J Bone Miner Res* 32: 1496-1504.
52. Pereira RF, O'Hara MD, Laptev AV, Halford KW, Pollard MD, et al. (1998) Marrow stromal cells as a source of progenitor cells for nonhematopoietic tissues in transgenic mice with a phenotype of osteogenesis imperfecta. *Proc Natl Acad Sci USA* 95: 1142-1147.
53. Guillot PV, Abass O, Bassett JH, Shefelbine SJ, Bou-Gharios G, et al. (2008) Intrauterine transplantation of human fetal mesenchymal stem cells from first-trimester blood repairs bone and reduces fractures in osteogenesis imperfecta mice. *Blood* 111: 1717-1725.
54. Chan JK, Gotherstrom C (2014) Prenatal transplantation of mesenchymal stem cells to treat osteogenesis imperfecta. *Front Pharmacol* 5: 223.
55. Gotherstrom C, Westgren M, Shaw SW, Aström E, Biswas A, et al. (2014) Pre- and postnatal transplantation of fetal mesenchymal stem cells in osteogenesis imperfecta: a two-center experience. *Stem Cells Transl Med* 3: 255-264.
56. Ault J, Sillence D (1996) A 20-year follow-up of osteogenesis imperfecta in Victoria, Australia. 6th International Conference on Osteogenesis Imperfecta. Zeist, The Netherlands.
57. Binder H, Conway A, Gerber LH (1993) Rehabilitation approaches to children with osteogenesis imperfecta: a ten-year experience. *Arch Phys Med Rehabil* 74: 386-390.
58. Binder H, Conway A, Hason S, Gerber L, Marini J, et al. (1993) Comprehensive rehabilitation of the child with osteogenesis imperfecta. *Am J Med Genet* 45: 265-269.
59. Tosi LL, Oetgen ME, Floor MK, Huber MB, Kennelly AM, et al. (2015) Initial report of the osteogenesis imperfecta adult natural history initiative. *Orphanet J Rare Dis* 10: 146.
60. Widmann RF, Laplaza FJ, Bitan FD, Brooks CE, Root L (2002) Quality of life in osteogenesis imperfecta. *Int Orthop* 26: 3-6.
61. Singer RB, Ogston SA, Paterson CR (2001) Mortality in various types of osteogenesis imperfecta. *J Insur Med* 33: 216-220.
62. McAllion SJ, Paterson CR (1996) Causes of death in osteogenesis imperfecta. *J Clin Pathol* 49: 627-630.